

## BLOOD KINETICS OF SULFAMONOMETHOXINE AND OXYTETRACYCLINE FOLLOWING INTRAUTERINE SPRAY INJECTION IN DAIRY COWS

C. GIRARDI\*†, G. RE\*, A. M. FARCA\*, M. DACASTO\*, E. FERRERO\* and M. EANDI†

\**Dipartimento di Patologia Animale, Università di Torino, via Nizza 52, 10126 Torino, Italia;* †*Istituto di Farmacologia e Terapia Sperimentale, Facoltà di Medicina e Chirurgia, Università di Torino, Italia*

*Received in final form 26 July 1989*

### SUMMARY

Intrauterine administration was performed in six Friesian cows with a disposable spray preparation containing 3 g of sulfamonomethoxine and 3 g of oxytetracycline, in order to investigate their serum kinetics. Sulfamonomethoxine levels were determined by a reversed-phase HPLC method, whilst oxytetracycline quantities were detected by a microbiological method.

The sulphonamide had a peak 1.17 h after the administration, the tetracycline reached its highest concentration after 8 h. The bioavailability of both drugs was low and detectable drug amounts were no longer recovered after 24 h.

KEY WORDS: sulfamonomethoxine, oxytetracycline, dairy cows, intrauterine administration, pharmacokinetics.

### INTRODUCTION

In recent years several antibacterial formulations have found wide applications in the intrauterine therapy of bacterial metritis in cows [1-3]. In veterinary practice, the spray preparations are nowadays considered the most suitable because of their easy handling and their good permeation of the uterine mucosa. Intrauterine administrations of antibacterials proved to be satisfactory, but despite this fact, they are considered as local treatments, although the absorption of many drugs from the uterus has been demonstrated [4-12].

The aim of the present report is to investigate the absorption and the related serum kinetics of a new combination of sulfamonomethoxine (SMM) and oxytetracycline (OTC) for intrauterine spray administration, since there are no data available in the literature referring to the passage of SMM from the uterus into the

†Correspondence to: Carlo Girardi, Cattedra di Farmacoterapia, Dipartimento di Patologia Animale, Facoltà di Medicina Veterinaria, via Nizza 52, 10126 Torino, Italia.

blood. A modified high-performance liquid chromatographic method [13] and a modified microbiological method [14] were used for the determination of SMM and OTC, respectively. The results of the kinetics of OTC are compared with those which we had obtained during previous experiments [15].

## MATERIALS AND METHODS

Six Friesian dairy cows, each weighing 450–500 kg, were used. During the trial, they were affected by endometritis clinically judged of moderate gravity. The cows were kept indoors and were fed with hay and tap water *ad libitum*.

Each cow received a dose of 3 g of sulfamonomethoxine and 3 g of oxytetracycline as a single intrauterine spray injection (Endospray preparations kindly supplied by IZO S.p.A. Brescia Italy). Blood samples were collected from the jugular vein 0, 0.5, 1, 2, 4, 8, 12, 24, 48 and 72 h after treatment. Samples were allowed to clot and then centrifuged to obtain sera which were stored at  $-20^{\circ}\text{C}$  until the analysis.

### *Sulfamonomethoxine determination*

A Spectra Physics 8100 high-performance liquid chromatograph, equipped with a SP 8840 UV/VIS detector and a SP 4290 printer/plotter integrator was used. A Merck stainless steel column (240 × 4 mm) packed with LiChrosorb RP 8, particle size 5  $\mu\text{m}$ , was connected with the chromatograph. An injection loop of 10  $\mu\text{l}$  was used.

The sulphonamide peaks, presenting a retention time of 3.5 min, were monitored at 260 nm; the detection limit was 0.02  $\mu\text{g/ml}$ . The method of external standard was employed.

The mobile phase was a mixture of phosphate buffer 0.067 M pH 6.7 and methanol (5:1). Samples were eluted isocratically at a flow rate of 1.6 ml/min and at a pressure of 115 atm. The sulfamonomethoxine sodium salt 99% pure was obtained from IZO S.p.A. Brescia, Italy (lot GL008 produced by Daiichi Seiyaku Co. Ltd. Tokyo Japan).

In order to make a calibration curve, five standard solutions were prepared at concentrations of 200, 20, 2, 0.2 and 0.02  $\mu\text{g SMM/ml}$  respectively (Fig. 1). The relation between SMM concentration and peak-height ratio (versus external standard) was linear ( $r=0.99$ ).

### *Sample preparation*

One ml of bovine serum was added to 4 ml of perchloric acid (0.33 M). The solution was mixed on a Vortex mixer and subsequently allowed to stand for 10 min. After centrifugation at 3000 rpm for 10 min and filtration through paper filter, the supernatant was ready for the injection into the column.

The precision and accuracy of the analytical method were evaluated by processing as replicates on different days, serum samples ( $n=4$ ) added with known standard amounts (0.2–200  $\mu\text{g SMM/ml}$ ), obtaining a recovery of  $80 \pm 7\%$  and a coefficient of variation ranging from 3.2% to 7.1%.

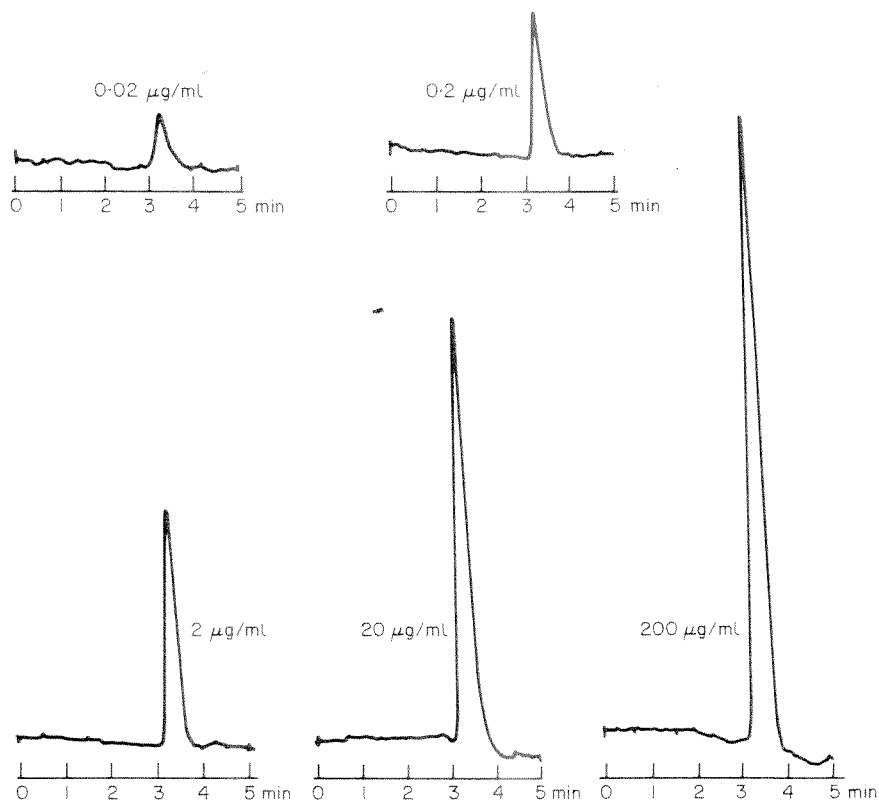


Fig. 1. Chromatograms of sulfamonomethoxine standard solutions.

#### *Oxytetracycline determination*

A microbiological hole-plated agar method, using *Bacillus cereus* ATCC 9634 as the assay organism, was performed. The sensitivity of this microbiological system was established to be  $0.1 \mu\text{g/ml}$ . Standard solutions ( $0.1$ – $5 \mu\text{g/ml}$ ) were prepared in purified water using oxytetracycline 99% pure (lot MP511470 Farmitalia Milan, Italy).

The calibration curve was obtained by plotting on semilogarithmic paper the concentration of the standard solutions on the y-axis and the diameter of the zone of inhibition on the x-axis and was linear ( $r > 0.99$ ). The concentrations of the unknown specimens were determined from the standard curve after measuring the zones of inhibition.

Cross-tests were performed on each method in order to reveal possible interferences during the assays.

#### *Pharmacokinetic parameters*

Kinetic parameters shown in Tables I and II were calculated by standard methods [16]. As regards each drug concentration against time curve, the

**Table I**  
Pharmacokinetic parameters of sulfamonomethoxine

Parameter	Cow						$\bar{x} \pm SD$
	A	B	C	D	E	F	
$C_{max}$ ( $\mu\text{g/ml}$ )	1.42	1.14	2.17	1.71	1.60	1.26	$1.55 \pm 0.37$
$T_{max}$ (h)	1.0	1.0	2.0	2.0	0.5	0.5	$1.17 \pm 0.68$
$K$ ( $\text{h}^{-1}$ )	0.36	0.28	0.31	0.18	0.22	0.32	$0.28 \pm 0.07$
$t_{1/2K}$ (h)	1.90	2.50	2.25	3.75	3.20	2.15	$2.63 \pm 0.71$
AUC ( $\mu\text{g/ml/g}$ )	6.83	7.18	9.70	8.81	6.04	3.76	$7.05 \pm 2.10$
0-8 h							
AUC ( $\mu\text{g/ml/h}$ )	7.12	8.39	10.76	11.86	7.95	4.33	$8.40 \pm 2.68$
0- $\infty$							

$C_{max}$ , highest plasma concentration;  $T_{max}$ , time at which  $C_{max}$  occurs (hours);  $K$ , first-order elimination rate constants;  $t_{1/2K}$ , half-time of the first-order elimination rate constant; AUC, total area under the plasma drug concentration versus time curve.

**Table II**  
Pharmacokinetic parameters of oxytetracycline

Parameter	Cow						$\bar{x} \pm SD$
	A	B	C	D	E	F	
$C_{max}$ ( $\mu\text{g/ml}$ )	0.97	1.35	1.59	0.70	0.97	1.59	$1.20 \pm 0.37$
$T_{max}$ (h)	8.0	8.0	8.0	8.0	8.0	8.0	$8.0 \pm 0.0$
$K$ ( $\text{h}^{-1}$ )	0.058	0.058	0.029	0.58	0.058	0.058	$0.05 \pm 0.01$
$t_{1/2K}$ (h)	12.0	12.0	24.0	12.0	12.0	12.0	$14.0 \pm 4.69$
AUC ( $\mu\text{g/ml/h}$ )	7.67	9.12	11.15	5.65	7.15	8.15	$8.15 \pm 1.87$
0-8 h							
AUC ( $\mu\text{g/ml/h}$ )	11.12	12.57	22.87	9.1	10.60	11.60	$12.98 \pm 4.98$
0- $\infty$							

$C_{max}$ , highest plasma concentration;  $T_{max}$ , time at which  $C_{max}$  occurs (h);  $K$ , first-order elimination rate constant;  $t_{1/2K}$ , half-time of the first-order elimination rate constant; AUC, total area under the plasma drug concentration versus time curve.

independent-model parameters  $K$ ,  $t_{1/2}$ , AUC 0-8 h, AUC 0- $\infty$ ,  $C_{max}$  and  $T_{max}$ , were evaluated. The  $K$  values were calculated on the tail of every single curve using a least squares regression analysis [16]. The areas under the curve were estimated by means of the trapezium method from  $T_0$  to  $T_n$  h and by the  $C_n/K$  integral from  $T_0$  to infinity.

## RESULTS AND DISCUSSION

Absorption of the drugs through the uterine mucosa, albeit in small amounts, occurred in all treated cows. The serum concentrations found during the experimental time are summarized in Table III and IV. The extent of absorption was calculated from areas under the curve.

Table III  
Sulfamonomethoxine serum levels ( $\mu\text{g/ml}$ )

	0	30 min	1 h	2 h	4 h	8 h	12 h	24 h	48 h	72 h
Cow A	—	1.24	1.42	1.38	0.96	0.10	—	—	—	—
Cow B	—	1.04	1.14	1.14	1.14	0.34	—	—	—	—
Cow C	—	1.27	1.94	2.17	0.88	0.33	—	—	—	—
Cow D	—	1.48	1.55	1.71	1.08	0.55	—	—	—	—
Cow E	—	1.60	1.47	1.26	0.75	0.28	—	—	—	—
Cow F	—	1.26	0.90	0.62	0.51	0.32	—	—	—	—
$\bar{x} \pm \text{SD}$		1.32 $\pm 0.20$	1.40 $\pm 0.36$	1.38 $\pm 0.53$	0.89 $\pm 0.29$	0.32 $\pm 0.14$				

—,  $< 0.02 \mu\text{g/ml}$ .

Table IV  
Oxytetracycline serum levels ( $\mu\text{g/ml}$ )

	0	30 min	1 h	2 h	4 h	8 h	12 h	24 h	48 h	72 h
Cow A	—	—	—	0.70	0.78	0.97	0.20	0.10	—	—
Cow B	—	—	—	0.55	0.83	1.35	0.20	0.10	—	—
Cow C	—	—	0.20	0.70	0.97	1.59	0.34	0.24	—	—
Cow D	—	—	0.20	0.51	0.51	0.70	0.20	0.10	—	—
Cow E	—	—	—	0.55	0.68	0.97	0.20	0.10	—	—
Cow F	—	—	0.20	0.36	0.70	1.59	0.20	0.10	—	—
$\bar{x} \pm \text{SD}$			0.10 $\pm 0.11$	0.56 $\pm 0.13$	0.75 $\pm 0.16$	1.20 $\pm 0.37$	0.22 $\pm 0.06$	0.12 $\pm 0.06$		

—,  $< 0.1 \mu\text{g/ml}$ .

The average concentrations in serum of SMM and OTC, plotted logarithmically against time are illustrated in Fig. 2. The results pointed out a rapid absorption of SMM from the uterine lumen, as detectable concentrations in serum were already present in the 0.30-h sample. The  $T_{\text{max}}$  was 1.17 h. In spite of this, OTC levels rose in the serum after 2 h presenting a  $T_{\text{max}}$  of 8 h, and confirming data reported in literature [10]. Moreover SMM presented a more rapid elimination phase ( $t_{1/2k}$  2.63 h) than OTC ( $t_{1/2k}$  14 h).

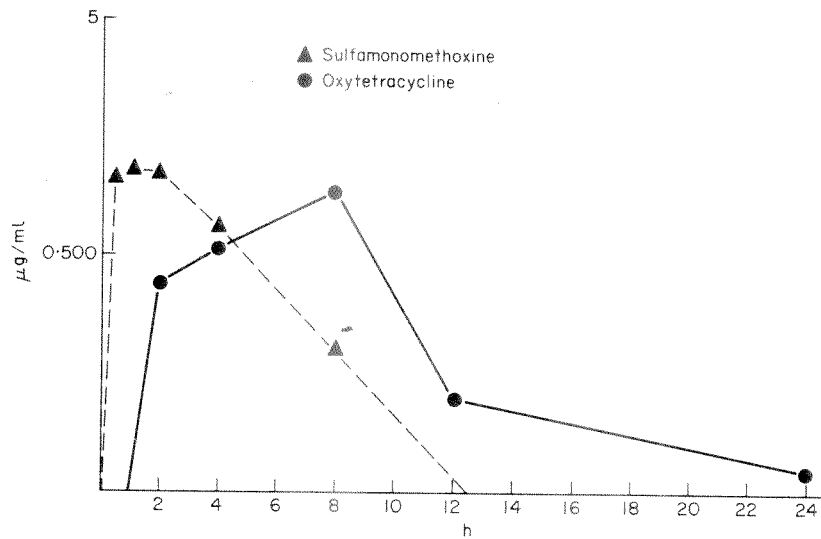


Fig. 2. SMM and OTC serum concentrations (mean values).

A metabolite peak, probably the  $N_4$ -acetylated sulphonamide, appeared close to the SMM peak showing a retention time of about 3 min (Fig. 3).

A comparison of the present results with those reported by other authors after i.v. and i.m. administration of SMM and OTC [7-20] emphasizes that the systemic

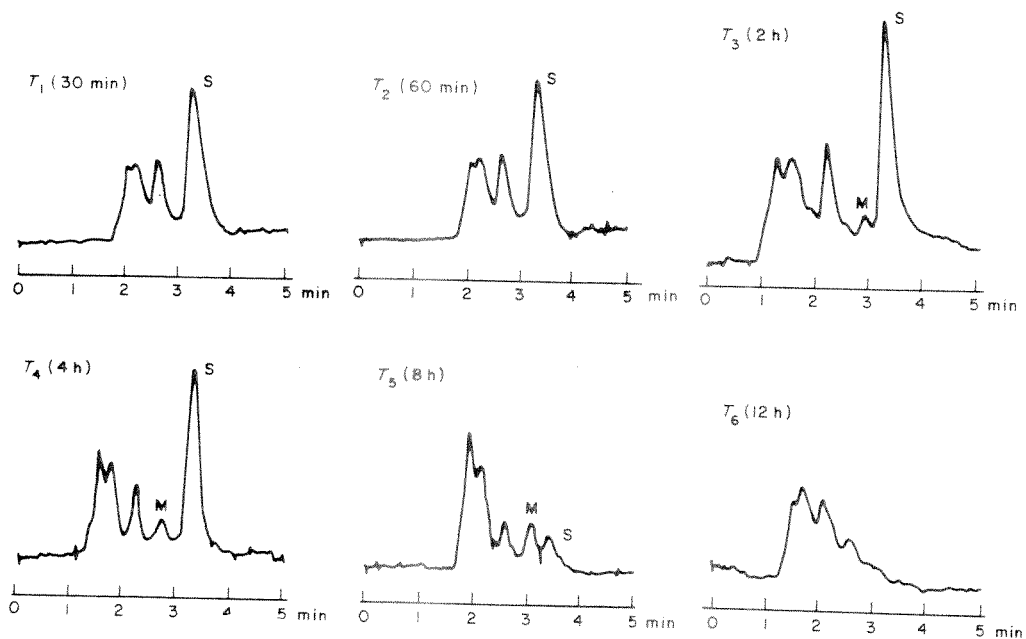


Fig. 3. Chromatograms of a cow treated with SMM: S, sulfamonomethoxine peak; M, metabolite peak.

bioavailability was low, as demonstrated by the values of  $C_{max}$  and AUC (Tables I and II). Our results are in close agreement with those already published [8, 10] upon the intrauterine administration of other antibacterials. As far as OTC is concerned, it is interesting to compare data resulting from the present study with those obtained by us in a previous investigation, during which dairy cows of the same herd were treated by intrauterine infusion with an equivalent dose of this drug, but using different kinds of vehicle [15].

In fact, no detectable serum levels of OTC were found when we used polyethylene glycol as the only vehicle for the infusion. By contrast, the uterine infusion of an aqueous solution of tetracycline gave serum levels almost resembling those achieved with OTC spray injection (Fig. 4).

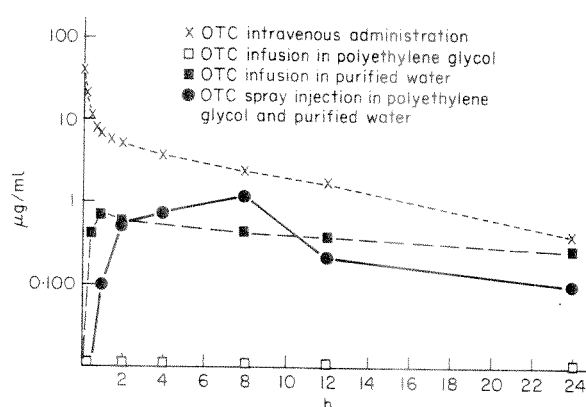


Fig. 4. Oxytetracycline, comparison between OTC spray injection and OTC infusion using different kinds of vehicle.

The slower absorption of sprayed OTC was probably due to the presence of glycol in the vehicle of the pharmaceutical preparation [21-23].

Finally, we suggest that the higher peak of OTC in serum could be ascribed to the spray form used and able to improve the endoperitoneal absorption through the uterine tubes [24] and the crossing of the uterine mucosa.

## REFERENCES

1. Oxender WD, Seguin BE. Bovine intrauterine therapy. *JAVMA* 1976; **168**: 217-19.
2. Seguin BE, Morrow DA, Oxender WD. Intrauterine therapy in the cow. *JAVMA* 1974; **164**: 609-12.
3. Signorini G, Ferrari A, Pera R, Gatti R, Negrelli AD. Valutazione della efficacia terapeutica nella vacca da latte di un antibiotico a lunga attività. *Ob Doc Vet* 1983; **9**: 67-8.
4. Ayliffe TR, Noakes DE. Intra-uterine absorption of sodium benzylpenicillin in the cow. *J Vet Pharmacol Ther* 1978; **1**: 267-71.
5. Black WD, Mackay AL, Doig PA, Claxton MJ. A study of drug residues in milk following intrauterine infusion of antibacterial drugs in lactating cows. *Can Vet J* 1979; **20**: 354-7.

6. Caudle AB, Brooks P, Frank JF. Measurement of ampicillin in milk after intrauterine infusion of dairy cows. *Vet Med* 1983; **78**: 239-42.
7. Haaland MA, Manspeaker JE, Moreland TW. Antibiotic residues in milk after intrauterine infusion. *Vet Med* 1984; **79**: 382-6.
8. Huang J, Hawkinson D, Miller GE. Passage of sulfadimethoxine from the bovine uterus into blood, milk and urine. *J Dairy Sci* 1972; **55**: 705.
9. Kaneene JB, Coe PH, Smith JH, *et al*. Drug residues in milk after intrauterine injection of oxytetracycline, lincomycin-spectinomycin and povidone-iodine in cows with metritis. *Am J Vet Res* 1986; **47**: 1363-5.
10. Miller GE, Bergt GP. Oxytetracycline in bovine plasma, milk, and urine after intrauterine administration. *J Dairy Sci* 1976; **59**: 315-17.
11. Miller GE, Rouse G. Passage of drugs from the bovine uterus into blood, milk and urine. *J Dairy Sci* 1970; **53**: 652.
12. Righter HF, Mercer HD, Kline AD, Carter GG. Absorption of antibacterial agents by the bovine involuting uterus. *Can Vet J* 1975; **16**: 10-15.
13. Vree TB, Hekster YA, Baars AM, Damsma JE, van der Kleijn E. Determination of trimethoprim and sulfamethoxazole (cotrimoxazole) in body fluids of man by means of high-performance liquid chromatography. *J Chromatogr* 1978; **146**: 103-12.
14. Siddique IH, Loken KI, Hoyt HH. Antibiotic residues in milk transferred from treated to untreated quarters in dairy cattle. *JAVMA* 1965; **146**: 589-93.
15. Girardi C, Farca AM, Eandi M, Leonori Cecina GP, Chiaretta G. Farmacocinetica e biodisponibilità sistemica di diverse preparazioni a base di ossitetraciclina somministrate per via endouterina in vacche da latte. *Atti Soc Ital Buiatria* 1988; **20**: 383-402.
16. Baggott JD. *Principles of drug disposition in domestic animals: the basis of veterinary clinical pharmacology*. Philadelphia, USA: W.B. Saunders Co., 1977.
17. Mevius DJ, Nouws JFM, Breukink HJ, Vree TB, Driessens F, Verkaik R. Comparative pharmacokinetics, bioavailability and renal clearance of five parenteral oxytetracycline-20% formulations in dairy cows. *Vet Quarterly* 1986; **8**: 285-94.
18. Nouws JFM, Breukink HJ, Binkhorst GJ, *et al*. Comparative pharmacokinetic and bioavailability of eight parenteral oxytetracycline-10% formulations in dairy cows. *Vet Quarterly* 1985; **7**: 306-14.
19. Nouws JFM, Van Ginneken CAM, Ziv G. Age-dependent pharmacokinetics of oxytetracycline in ruminants. *J Vet Pharmacol Ther* 1983; **6**: 59-66.
20. Nouws JFM, Vree TB, Termond E, *et al*. Pharmacokinetics and renal clearance of oxytetracycline after intravenous and intramuscular administration to dairy cows. *Vet Quarterly* 1985; **7**: 296-305.
21. Al-Guedawy SA. Effect of vehicle on intrauterine absorption of gentamicin in cattle. *Theriogenology* 1983; **19**: 771-8.
22. Gobbi L, Signorini GC, Ballarini G, Morselli P. Trattamento endouterino con cefalessina: valutazioni dell'effetto di tre diversi veicoli. *Atti Soc Ital Buiatria* 1985; **17**: 537-48.
23. Malvisi Stracciari J, Zaghini A. L'influenza del glicole propilenico e del polivinilpirrolidone sul comportamento dei farmaci iniettabili. *Ob Doc Vet* 1982; **11**: 9-15.
24. Ayliffe TR, Noakes DE. Effect of ligation of the bovine uterine tubes on absorption of intrauterine infusions of sodium benzylpenicillin. *Vet Rec* 1986; **118**: 243-4.